

National Library



	ot Med	icine		4 1 4 1 1 1 5	14
PubMed	Nucleotide	Protein	Genome	Structure	PopSet
Search PubMed	₹ for			Go	Clear
	Limits	Preview/Inde	x History	Clipboard	
Entrez PubMed	Display Abs	tract S	ave Text Ord	er Add to Clip	bboard
	1: Adv Exp	Med Biol 1992;32	21:1-5	Relate	d Articles, Books
PubMed Services	Pancreatic islet cell regeneration and growth. Introduction.				
	Vinik AI				
Related Resources	Eastern Virginia Medical School, Diabetes Institutes, Norfolk 23510.				
		on Types:			•
		view view, tutorial			
	PMID: 1449076, UI: 93080098				
	•				
	Display Abs	tract 🔽 S	ave Text Ord	er Add to Clip	board

Write to the Help Desk NCBI | NLM | NIH Department of Health & Human Services Freedom of Information Act | Disclaimer

INTRODUCTION

Aaron I. Vinik, M.D., Ph.D.1

¹Eastern Virginia Medical School The Diabetes Institutes Norfolk, Virginia 23510

This symposium, held in June 1991, was a gathering of international scientists to exchange their views on current concepts of cell growth and differentiation. Each scientist was asked to present a topic of their research related to cell growth and regeneration and to participate in a round table conference elaborating on current knowledge and sharing their experiences. By furthering this promising area of endeavor, a means of understanding ontogeny of cell development and of providing insights into tumor biology would prevail. Of prime importance was the anticipation that new information from a better understanding of the normal evolution of the pancreatic islet would generate alternative approaches to curing diabetes. This forward serves as a short introduction to the concept of pancreatic islet regeneration and the models currently in use to study the process.

DEVELOPMENTAL ORIGIN OF ISLETS DURING EMRYOGENESIS

The developing pancreas appears as a protrusion from the dorsal surface of the embryonic gut. The different islet cell types appear sequentially during development in vivo. It therefore seems reasonable to propose that coordinated growth is dependent upon specificity of growth factors.

Islet cells in vivo also express several neuroectodermal antigens, for example, PGP 9.5,2 neurone-specific enolase (NSE),3 synaptophysin,4 A2B5,5 phenylethanolamine N-methyltransferase (PNMT) and aromatic amino acid decarboxylase (AADC).6 The endocrine cells of the GEP axis are capable of amine precursor uptake and decarboxylation and have, therefore, been given the actonym APUD.7 The morphologic similarity of APUD cells suggested a common embryologic origin, which was believed to be the neural crest but later revised to include the neuroectoderm, or in the case of some of the endocrine cells, from the dorsal placoderm.

Studies by Ledouarin, 8 Pictet, 9 Andrew, 10 and their coworkers have cast doubt on this hypothesis, and most workers agree that these cells should be classified according to their secretory products, i.e., gastrin, somatostatin, glucagon. PP, etc. However, it is now thought that β -cells do not have APUD characteristics and are likely to be derived from gut, but express neuronal antigens such as the catecholamine biosynthetic enzyme tyrosine hydroxylase (TH). 11 The generally held belief that the neuronal characteristics of these cells indicated an ectodermal origin during mammalian embryogenesis has largely been dispelled.

During development in vivo, the phenotype of the mature islet cells appear sequentially. β-cells arise from progenitor cells localized in the pancreatic duct and these precursors transiently express TH while migrating away from the duct to populate a new islet. This suggests that the pancreatic duct is a source of endocrine stem cells throughout embryogenesis without the need to postulate a neuroendocrine origin. This notion is supported by the finding that the embryonic pancreatic duct in vitro is able to regenerate a new pancreas containing exocrine and endocrine cells expressing only peptides (mature cells), and cells containing both TH and a hormone (immature cells). 12,13

Teitelman has shown that pancreatic cells of endocrine origin can indeed express several neuronal antigens in addition to the peptide hormones. 11 She further showed that in the mouse embryo a primitive undifferentiated cell(s) led to sequential appearance of at least 4 different cell types containing either a hormone (e.g., glucagon), a catecholamine enzyme (tyrosine hydroxylase, TH) or combinations of these. Under appropriate conditions these cells can be shown to differentiate into either neurites or adult endocrine cells. During regeneration, expression of neural antigens by developing cells was found to constitute an early phase to be replaced by the adult hormone secreting counterpart. Rosenberg and Vinik 14 have utilized a model for nesidioblastosis and shown that pancreatic ductal cells are capable of differentiating upon stimulation into adult endocrine cells capable of secreting insulin in a fully regulated manner.

ISLET CELL GROWTH AND DIFFERENTIATION

Factors which control the growth and functional maturation and differentiation of the human endocrine pancreas and gut during the fetal and post-natal periods are incompletely The role of the fetal mesenchyme in epithelial cell development and differentiation appears to be important. 15-17 Possible mechanisms of action include: (i) secretion of an inducing or transforming hormonal growth factor, (ii) information exchange through cell-to-cell contact via paracrine and juxtacrine actions of locally elaborated growth factors, and (iii) production of an extracellular matrix rich in growth promoting factors. The soluble peptide growth factors are trophic substances that regulate both cell proliferation and

differentiation and may be linked to islet growth.

One family of growth factors that may be implicated in islet growth are the somatomedins and their binding proteins. Insulin-like growth factors (IGFs) are important mediators of fetal and postnatal growth. Whereas these growth factors circulate (attached to binding proteins) they also act locally. Fetal rat islets release both IGF-I and IGF-II in vitro which may contribute to growth hormone-induced DNA synthesis. 18-21 It is apparent that the role of IGF-I, and the binding proteins, especially in the adult pancreas is far from clear. In this symposium Drs. LeRoith and Lauterio discuss aspects of IGF physiology, and LeRoith focuses upon differences in the role of the IGF's in regeneration of adult and fetal tissues. Dr. Hill emphasizes the role of the IGF's and their binding proteins in islet regeneration in the fetal pancreas at a time when the pancreas is susceptible to the influence of this particular group of growth factors. Dr. Bonner Weir also points out that in their model of islet regeneration after 90% pancreatectomy there is enhanced IGF gene expression in the ductules and certain connective tissue cells in contrast with the normal expression in capillary endothelial cells suggesting that IGFs may participate in the regeneration process after pancreatectomy in the rat.²² Dr. Nielsen, however, contests the suggestion that the IGF's are important as pancreatic trophic factors and based upon his observations growth hormone itelf may be pertinent, at least in pregnancy, a state in which islet hypertrophy is

Several important glycoprotein components of extracellular matrix - the "integrins"have also been recognized as playing a role in cell growth and differentiation. For example, fibronectin, laminin and tenascin. 23,24 Dr. Le Beau elaborates upon the role the integrins may play in cell regeneration. The role of these factors in the maintenance and replacement

of a functional islet cell mass in the adult pancreas remains to be determined.

ISLET CELL PROLIFERATION IN THE POST-NATAL PERIOD

Several models designed to induce exocrine and endocrine pancreatic regeneration have been developed. In the model developed by Bonner-Weir and colleagues, 22,25 there is regeneration of both exocrine and endocrine tissue following a 90% pancreatectomy in which the increase in β -cell mass occurs as a result of the replication of existing β -cells and not necessarily because of a process of new islet formation.²² This group have reported increased IGF-I mRNA production by capillary endothelial cells and proliferating ductules which may contribute to both endocrine and exocrine pancreas regeneration, but the precise role of IGFs needs to be has not been excluded. 22

Terazono et al deve treated with either n aurothioglucose resulting a substance termed reg p gene.28 Human reg mR levels in gastric mucosa expressed ectopically in transformed, proliferative reg is expressed in acina growth factor. Dr. Ol regeneration, elaborates this growth factor in isle and Rafaeloff.

Expression of an h tissue.30 The signification regeneration studied by over a period of weeks to

A second version of colleagues,31 deals with Hospital (NEDH) rats demonstrate that tumor with a profound reduction that results in rapid BmRNA levels. Whether high levels of reg mR ongoing at a low, consti this gene product.

Another model has is observed in transgen These mice, in which d duct cell proliferation as along the apical region indicating that diabetes destructive process. T induce regeneration may

In 1982, we develop 8 weeks of age are fertil of cell proliferation that hamster pancreatic duct was observed. The m proliferation and differ processes were shown t an extract prepared from other hamsters, but this pancreas was administe developments in the us called flotropin.

The presence of th been hypothesized but reviews the current kn carried out to further de the release of a variety and differentiation of d with factors important i possible mechanism of also discussed.

in can indeed express several the further showed that in the ntial appearance of at least 4 m), a catecholamine enzyme appropriate conditions these ult endocrine cells. During Is was found to constitute an counterpart. Rosenberg and that pancreatic ductal cells are ne cells capable of secreting

ion and differentiation of the atal periods are incompletely elial cell development and misms of action include: (i) or, (ii) information exchange of locally elaborated growth towth promoting factors. The ate both cell proliferation and

ed in islet growth are the factors (IGFs) are important 1 factors circulate (attached to oth IGF-I and IGF-II in vitro nesis. 18-21 It is apparent that lult pancreas is far from clear. ects of IGF physiology, and egeneration of adult and fetal ir binding proteins in islet is susceptible to the influence also points out that in their inhanced IGF gene expression with the normal expression in te in the regeneration process ntests the suggestion that the ipon his observations growth in which islet hypertrophy is

ular matrix - the "integrins"-differentiation. For example, es upon the role the integrins maintenance and replacement æ determined.

PERIOD

e pancreatic regeneration have and colleagues, 22,25 there is ng a 90% pancreatectomy in lication of existing β-cells and 22 This group have reported ells and proliferating ductules s regeneration, but the precise role of IGFs needs to be elucidated and the possibility that other growth factors participate

has not been excluded. 22

Terazono et al developed a model in which 90% pancreatectomized rats or mice are treated with either nicotinamide, (a poly ADP-ribose synthetase inhibitor), or aurothioglucose resulting in exocrine and endocrine cell regeneration, and the appearance of a substance termed reg protein. 26.27 The gene encoding this protein has been termed the reg gene. 28 Human reg mRNA has been detected predominantly in the pancreas, and at lower levels in gastric mucosa and in the kidney. 29 Reg gene protein has also been found to be expressed ectopically in colon and rectal tumors, 29 linking enhanced reg expression to the transformed, proliferative state, at least for some cell types. Current evidence suggests that reg is expressed in acinar tissue and not regenerating islets. It may therefore be a paracrine growth factor. Dr. Okamoto, who pioneered the work on Reg gene and pancreatic regeneration, elaborates here upon his new findings. The controversy over the relevance of this growth factor in islet regeneration in other models is further discussed by Drs. Newgard and Rafaeloff.

Expression of an homologous gene, termed rig, has been identified in insulinoma tissue. 30 The significance of these genes remains to be determined. In both models of regeneration studied by Okamoto and colleagues, 26,27,29 islet size increased above normal

over a period of weeks to months. A second version of the 90% pancreatectomized rat model studied by Newgard and colleagues, 31 deals with reg expression in insulinoma-bearing New England Deaconess Hospital (NEDH) rats relative to normal controls and following tumor resection. They demonstrate that tumor implantation causes a sharp reduction in reg expression associated with a profound reduction in non-tumor islet size and that removal of the tumor, a maneuver that results in rapid β -cell proliferation, results in a large but transient induction in reg mRNA levels. Whether reg protein is β -cytotrophic is still an open question. The fact that high levels of reg mRNA are present in normal animals, in which β -cell replication is ongoing at a low, constitutive rate, are seemingly at odds with a growth-promoting role for this gene product.

Another model has been suggested by Dr. Sarvetnick³² in which a regenerative process is observed in transgenic mice expressing Interferon-gamma in their pancreatic β -cells. These mice, in which diabetes ensues following immunodestruction of their β -cells, show duct cell proliferation and the appearance of more primitive neuroendocrine progenitor cells along the apical regions of the ducts. Dr. Sarvetnick discusses her most recent findings indicating that diabetes may not develop provided that the regenerative process outstrips the destructive process. This, we believe, is an important principle whereby an approach to induce regeneration may not be unreasonable in our quest for a cure for diabetes.

In 1982, we developed a unique model for islet regeneration in hamsters, 33,34 Hamsters 8 weeks of age are fertile, considered to be adult animals and respond better to the induction of cell proliferation than do older animals. 33,34 By producing partial obstruction of the hamster pancreatic duct by cellophane wrapping, new islet formation from ductal elements was observed. The mechanism by which partial obstruction in our model induces cell proliferation and differentiation is unknown. Using a parabiotic experimental design, these processes were shown to be mediated by paracrine and/or autocrine mechanisms. 30 Indeed, an extract prepared from a wrapped pancreas exhibited trophic activity when injected into other hamsters, but this was not observed when an extract prepared from a non-wrapped pancreas was administered. Drs. Rosenberg and Vinik review the data and the more recent developments in the use of a cytosol extract, containing the growth factor which we have called llotropin.

The presence of this specific growth factor, Ilotropin, in the β -cell cytosol extract has been hypothesized but the identity of the peptide had not been established. Dr. Pittenger reviews the current knowledge of the nature of the growth factor and the studies he has carried out to further define its characteristics. However, cellophane wrapping may initiate the release of a variety of growth factors that contribute to the coordinated, timely growth and differentiation of ductal cells. These possibilities and other factors that may be shared with factors important in the nervous system are highlighted by members of the faculty. The possible mechanism of action of the various factors as well as therapeutic applications are

also discussed.

GROWTH FACTOR(S) AND NEOPLASIA

A great deal of interest is now being focused upon the factors responsible for initiation of growth, increase in cell number and size, differentiation into adult endocrine cells, growth cessation and cell maintenance. 36,37 The coincidental findings that the multiple endocrine neoplasia, type 1 syndrome (MEN-1) (combined occurrence of tum rs f the pituitary, pancreas and parathyroid glands) is associated with the loss of alleles on chromosome 11,38,39 the same chromosome on which the insulin gene has been located;40 the finding of parathyroid mitogenic activity in the plasma of patients with MEN-1;36,37 and evidence that patients with MEN-1 might also secrete mitogenic factors for pancreatic islet-cells into plasma,41 suggests a role for genetically determined circulating growth factors in the 'growth initiation' of these tumors.

It is apparent, therefore, that the conference was timeous and that the information pertinent to our appreciation of cell growth and differentiation is relevant to a possible cure for diabetes as well as a clearer understanding of factors that may be involved in pancreatic tumor formation. Conceptually, these conditions represent the two ends of a spectrum diabetes with endocrine cell failure and endocrine neoplasia with unbridled cell growth.

REFERENCES

- R.L. Pictet and J.W. Rutter, Development of the embryonic endocrine pancreas, in: "Handbook of Physiology," D.F. Steiner and M. Frenkel, eds., Washington D.C., American Physiological Society
- K.J. Thompson, J.F. Doran, P. Jackson, A.P. Dhillon, and J. Rode, PGP 9.5: a new marker for vertebrate neurons and endocrine cells, Brain Res. 278:224-28 (1983).
- 3. Polak, et al. Neuron-specific enolase, a marker for neuro-endocrine cells, in: Evolution and Tumor Pathology of the Neuro-endocrine System. Elsevier, New York (1984).
- 4. B. Weidenmann, W.W. Franke, C. Kuhn, R. Moll, and V.E. Gould, Synaptophysin: a marker protein for neuro-endocrine cells and neoplasms, Proc Natl Acad Sci USA, 83:3500-04 (1986).
- 5. G.S. Eisenbarth, K. Shimizu, M.A. Bowring, and S. Wells, Expression of receptors for fetanus toxin and monoclonal antibody A₂B₂ by pancreatic islet cells, Proc Natl Acad Sci USA. 79:5066-70 (1982).
- G. Teitelman, T.H. Joh, and D.J. Reis, Transformation of catecholaminergic precursor into glucagon (A) cells in mouse embryonic pancreas, Proc Natl Acad Sci USA. 78:5225-29 (1991).
- A.G.E. Pearse, Common cytochemical and ultrastructural characteristics of cells producing polypeptide hormones (the APUD series) and their relevance to thyroid and ultimobronchial C cells and calcitonin, Proc R Soc Lond (Biol). 170:71 (1968).

- N.M. LeDouarian and M.A. Teillet, The migration of neural crest cells to the wall of the digestive tract in avian embryo, J Embryol Exp Morphol. 30:31 (1973).
 R.L. Pictet, L.B. Rall, P. Phelps, and W.J. Rutter, The neural crest and the origin of the insulin-producing and other gastrointestinal hormone-producing cells, Science. 191:191 (1967).
 A. Andrew, An experimental investigation into the possible neural crest origin of pancreatic APUD (islet) cells, J Embryol Exp Morphol. 35:577 (1976).
- 11. G. Teitelman and J.K. Lee, Cell lineage analysis of pancreatic islet cell development: Glucagon and insulin cells arise from catecholaminergic precursor present in the pancreatic duct, Devel Biol. 121:454-56 (1987).
- G. Teitelman, J. Lee, and D.J. Reis, Differentiation of prospective mouse pancreatic islet cells during development in vitro and during regeneration, Dev Biol. 120:425-33 (1987).
- 13. R.W. Dudek and I.E. Lawrence, Morphologic evidence of interaction between adult ductal epithelium of pancreas and fetal foregut mesenchyme, Diabetes. 37:891-900 (1988).
- L. Rosenberg, W.P. Duguid, and A.I. Vinik, Cell proliferation in the pancreas of the Syrian golden hamster, Dig Dis Sci. 32:1185 (1987).
- R.L. Pictet, L. Rail, M. de Gasparo, and W.J. Rutter, Regulation of differentiation of endocrine cells during pancreatic development in-vitro, in: "Early diabetes in early life," R.A. Camerini-Davalos and H.S. Cole, eds., Academic Press, New York (1975).
- B.S. Spooner, H.I. Cohen, and J. Faubion, Development of the embryonic mammalian pancreas: The relationship between morphogenesis and cytodifferentiation, Dev Biol. 61:119 (1977).
- 17. R. Montesano, P. Mouron, M. Amherdt, and L. Orci, Collagen matrix promotes reorganization of pancreatic endocrine cell monolayers into islet-like organoids, J Cell Biol. 97:935 (1983).
- 18. I. Sweene and D.J. Hill, Growth hormone regulation of DNA replication, but not insulin production, is partly mediated by somatomedin C/insulin-like growth factor-I in isolated pancreatic islets from adult rats, Diabetologia. 32:191-97 (1989).
- 19. D.J. Hill, A. Frazer, I. Swe, P.K. Wirdnam, and R.D.G. Milner, Somatostatin-C in human fetal pancreas, Diabetes. 36:465 (1987).

- 20. A. Rabinovitch, C. Quigl activity (an insulin-li monolayer cultures.
- 21. J.M. Bryson, B.E. Tuch. pancreas in culture, J
- 22. J.S. Brockenbrough, G.C. 90% pancreatectomy
- 23. M.J. Politis, Exogeneous Reconstr Surg. 83:22
- 24. R. Chiquet-Ehrismann, E protein involved in ti (1986).
- 25. F. Smith, K. Rosen, L. Vil is localized to capilla
- 26. K. Terazono, H. Yamamo gene activated in regi
- 27. K. Terazono, Y. Uchiyam Expression of reg pre secretory granules, D
- 28. T. Watanabe, H. Yonekur of human reg gene as (1990).
- 29. C. Miyaura, et al, Express cell mass, Mol Endoc
- 30. S. Takasawa, K. Yamamo Diabetes. 35:1178 (19
- 31. L. Chen, M. Appel, T. Ala Regulating Islet Rege
- 32. N. Sarvetnick, Islet cell de (abstract) (1991).
- 33. L. Rosenberg, W.P. Dugus diabetes in the Syrian
- 34. L. Rosenberg and A.I. Vir paracrine and/or auto
- 35. L. Rosenberg and A.I. Vie from the "wrapped" p 36. M.L. Brandi, G.D. Aurba with familial multiple
- 37. S.J. Marx, K. Sakaguchi, in plasma from mem!
- Endocrinol Metab. 67 38. C. Larsson, B. Skogseid, 1 gene maps to chron 39. R.V. Thakker, P. Bouloux
- O'Riordan, Association alleles on chromoson 40. D. Owerbach, G.I. Bell, V
- arm of chromosome
- 41. M.K. McLeod, A.M. Tute mitogenic factor in pa November 15-18, 198

ctors responsible for initiation adult endocrine cells, growth igs that the multiple endocrine e of tumors of the pituitary, ss of alleles on chromosome been located;40 the finding of MEN-1:36,37 and evidence that for pancreatic islet-cells into g growth factors in the 'growth

ous and that the information n is relevant to a possible cure may be involved in pancreatic the two ends of a spectrum ith unbridled cell growth.

pancreas, in: "Handbook of , American Physiological Society

P 9.5: a new marker for vertebrate

ills, in: Evolution and Tumor 1984). naptophysin: a marker protein for 3:3500-04 (1986). of receptors for fetanus toxin and ad Sci USA. 79:5066-70 (1982). nergic precursor into glucagon (A) 5225-29 (1991). cs of cells producing polypeptide timobronchial C cells and calcitonin,

to the wall of the digestive tract in

the origin of the insulin-producing 191 (1967). st origin of pancreatic APUD (islet)

I development: Glucagon and pancreatic duct, Devel Biol.

use pancreatic islet cells during -33 (1987). between adult ductal epithelium of 388). ancreas of the Syrian golden

ferentiation of endocrine cells during R.A. Camerini-Davalos and H.S.

mic mammalian pancreas: The / Biol. 61:119 (1977). promotes reorganization of žell Biol. 97:935 (1983). on, but not insulin production, is isolated pancreatic islets from adult

tostatin-C in human fetal pancreas,

- 20. A. Rabinovitch, C. Quigley, T. Russell, Y. Patel, and D.H. Mintz, Insulin and multiplication stimulating activity (an insulin-like growth factor) stimulate islet β-cell replication in neonatal rat pancreatic monolayer cultures, Diabetes, 31:160 (1982).
- J.M. Bryson, B.E. Tuch, and R.C. Baxter, Production of insulin-like growth factor-II by human fetal pancreas in culture, J Endocrinol. 121:367-73 (1989).
- J.S. Brockenbrough, G.C. Weir, and S. Bonner-Weir, Discordance of exocrine and endocrine growth after 90% pancreatectomy in rats, Diabetes. 37:232-36 (1988).
- 23. M.J. Politis, Exogeneous laminin induces regenerative changes in traumatized sciatic and optic nerve, Plas Reconstr Surg. 83:228-35 (1989).
- 24. R. Chiquet-Ehrismann, E.J. Mackie, C.A. Pearson, and T. Sakakura, Tenascin: an extracellular matrix protein involved in tissue interactions during fetal development and oncogenesis, Cell. 47:131-39 (1986).
- 25. F. Smith, K. Rosen, L. Villa-Kamoroff, et al, Enhanced IGG-I gene expression in regenerating rat pancreas is localized to capillaries and proliferating ductules, Diabetes. 39(Suppl 1):66A (1990).
- K. Terazono, H. Yamamoto, S. Takasawa, K. Shiga, Y. Yonemura, Y. Tochino, and H. Okamoto, A novel gene activated in regenerating islets, J Biol Chem. 262:2111 (1988).
- K. Terazono, Y. Uchiyama, M. Ide, T. Watanabe, H. Yonekura, H. Yamamoto, and H. Okamoto, Expression of reg protein in rat regenerating islets and its co-localization with insulin in beta cell secretory granules, Diabetologia. 33:1 (1990).
- 28. T. Watanabe, H. Yonekura, K. Terazono, H. Yamamoto, and H. Okamoto, Complete nucleotide sequence of human reg gene and its expression in normal and tumoral tissues, J Biol Chem. 265:7432-39
- 29. C. Miyaura, et al, Expression of reg/PSP, a pancreatic exocrine gene: Relationship to changes in islet β cell mass, Mol Endocrinol (1991, in press).
- 30. S. Takasawa, K. Yamamoto, K. Terazono, and H. Okamoto, Novel gene activated in rat insulinoma, Diabetes. 35:1178 (1986).
- 31. L. Chen, M. Appel, T. Alam, C. Miyaura, A. Sestak, J. O'Neil, R. Unger, and C. Newgard, Factors Regulating Islet Regeneration in the Post-Insulinoma NEDH Rat.
- 32. N. Sarvetnick, Islet cell destruction and regeneration in IFN-y transgenic mice, J Cell Biochem. CB019:49 (abstract) (1991).
- 33. L. Rosenberg, W.P. Duguid, R.A. Brown, and A.I. Vinik, Induction of islet cell proliferation will reverse diabetes in the Syrian golden hamster, Diabetes. 37:334 (1988).
- L. Rosenberg and A.I. Vinik, Regulation of pancreatic islet growth and differentiation: Evidence for paracrine and/or autocrine growth factor(s), Clin Res. 38:271A (1990).

- L. Rosenberg and A.I. Vinik, In vitro stimulation of hamster pancreatic duct growth by an extract derived from the "wrapped" pancreas, (submitted for publication).
 M.L. Brandi, G.D. Aurbach, L.A. Fitzpatrick, et al, Parathyroid mitogenic activity in plasma from patients with familial multiple endocrine neoplasia type 1, N Engl J Med. 314:1287-93 (1986).
 S.J. Marx, K. Sakaguchi, J. Green, G.D. Aurbach, and M.L. Brandi, Mitogenic activity on parathyroid cells in plasma from members of a large kindred with multiple endocrine neoplasia Type 1 J Clin S.J. Marx, K. Sakagueni, J. Green, G.D. Auroach, and M.L. Brandi, Mulogenic activity on paramytoid cells in plasma from members of a large kindred with multiple endocrine neoplasia Type 1, J Clin Endocrinol Metab. 67:149-53 (1988).
 C. Larsson, B. Skogseid, K. Oberg, Y. Nakamura, and M. Nordenskjold, Multiple endocrine neoplasia type 1 gene maps to chromosome 11 and is lost in insulinoma, Nature. 332:85-87 (1988).
 R.V. Thakker, P. Bouloux, C. Wooding, K. Chotai, P.M. Broad, N.K. Spurr, G.M. Besser, and J.L. O'Riordan. Association of parathyroid tumors in multiple endocrine neoplasia type 1 with loss of
- N. V. Inakker, F. Bouloux, C. Wooding, K. Chotal, F.M. Broad, N.K. Spurr, G.M. Besser, and J.L. O'Riordan, Association of parathyroid tumors in multiple endocrine neoplasia type 1 with loss of alleles on chromosome 11, N Engl J Med. 321:218-24 (1989).
 D. Owerbach, G.I. Bell, W.J. Rutter, J.A. Brown, and T.B. Shows, The insulin gene is activated on the short arm of chromosome 11 in humans, Diabetes. 30:267-70 (1981).
 M.K. McLeod, A.M. Tutera, N.W. Thompson, and A.I. Vinik, Evidence for a pancreatic islet-cell mitogenic factor in natients with MEN-1. Association for Academic Surgery. Louisville. Kentucky.
- mitogenic factor in patients with MEN-1, Association for Academic Surgery, Louisville, Kentucky, November 15-18, 1989.